

Activation of Human CD4⁺CD25⁺CD127^{lo}Treg with Alloantigen and rIL-2 Induces IFNGR

Al-Atiyah R*, Verma ND*, Kawalia P, Tran GT, Hodgkinson S.J, Hall BM



Immune Tolerance Laboratory, Faculty of Medicine, South Western Sydney Clinical School, UNSW Australia and Ingham Institute, Liverpool Hospital, Liverpool, NSW Australia



INTRODUCTION

Activation of human alloantigen specific CD4⁺CD25⁺CD127^{lo}Foxp3⁺Treg that induce tolerance is a key goal. In rats, we have shown that CD4⁺CD25⁺Treg activated with alloantigen and rIL-2 express more Foxp3 and CD25 and receptors for IFN-γ (IFNGR) and IL-12 (IL-12Rβ2). These cells are more potent suppressors of specific alloactivation (Verma et al, 2009).

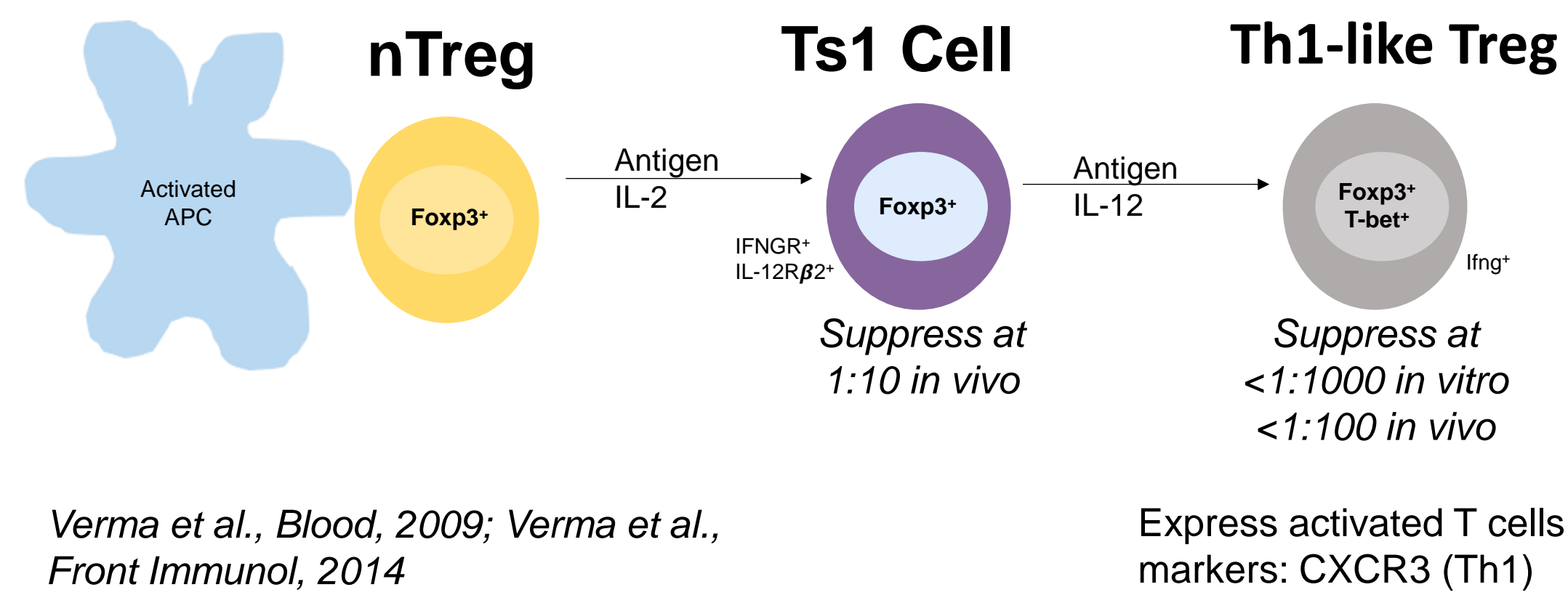
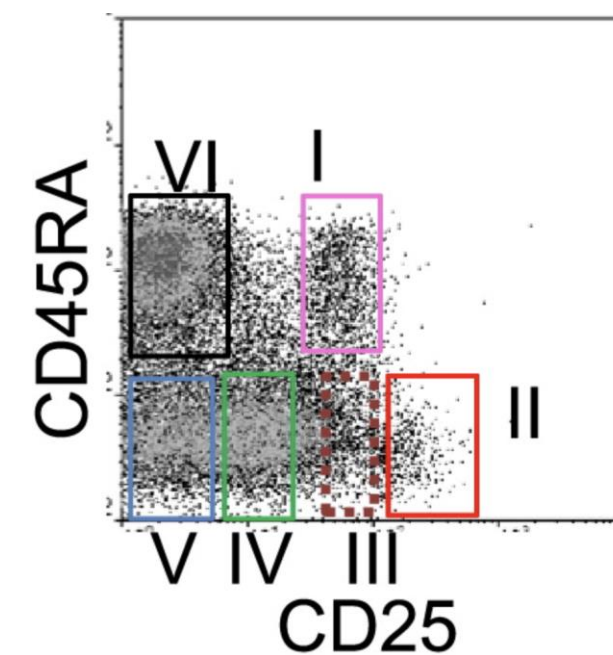


Figure 1. Proposed pathway for activation/proliferation of alloantigen-specific nTreg in rats. Initial culture of Foxp3⁺ nTreg and alloantigen and IL-2 gives rise to Ts1 cells that can suppress T effector cells (Teff) at a ratio of 1:10 Ts1:Teff in vivo. Reculturing Ts1 cells with alloantigen and IL-12 induces Th1-like Treg that have further enhanced suppressive capacity.

Human CD4⁺ cells based on CD45RA and Foxp3/CD25 are shown to comprise of 5 populations (Figure 2); Pop I (Foxp3⁺CD25⁺CD45RA⁺) as resting Treg; Pop II (Foxp3^{hi}CD25^{hi}CD45RA⁻) as activated Treg and Pop III (Foxp3⁺CD25⁺CD45RA⁻) including both Treg and activated effector T cells (Miyara, 2009). Pop II and III express chemokine receptors of activated T cells, including CXCR3 (Th1), CCR6 (Th17), CCR4 (Th2) which promote migration to inflammatory site.

Here, we examined activation of human CD4⁺CD25⁺CD127^{lo}Treg and subpopulations and studied their FACS profile for Treg markers, cytokine receptor and chemokine receptors.

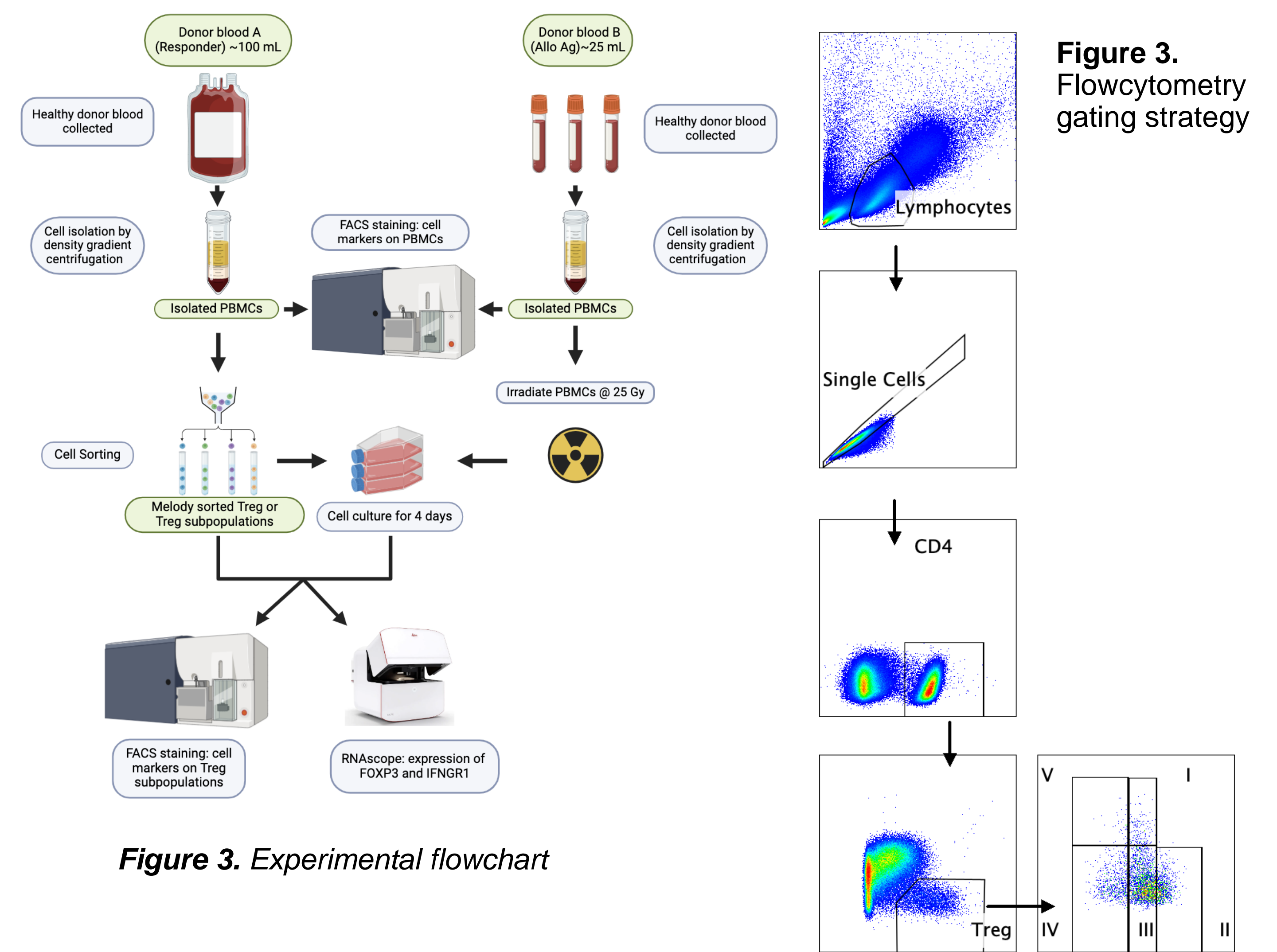
Figure 2: CD4⁺ populations



METHODS

CD4⁺CD127^{lo}CD25⁺Treg isolated by FACS from blood of healthy volunteers were cultured for 4 days with rIL-2 and irradiated allostimulators (alloS). Cells were examined using multicolour flow cytometry. Data was acquired on BD FACSCanto II using BD FACSDiva software (v8.0) and analysed using FlowJo for shifts in Pop I, II, III and chemokine receptors.

Some cells were stained for FOXP3 and IFNGR1 using RNAscope.



RESULTS

Culture with rIL-2 and alloantigen increases proportion of Pop II within Treg

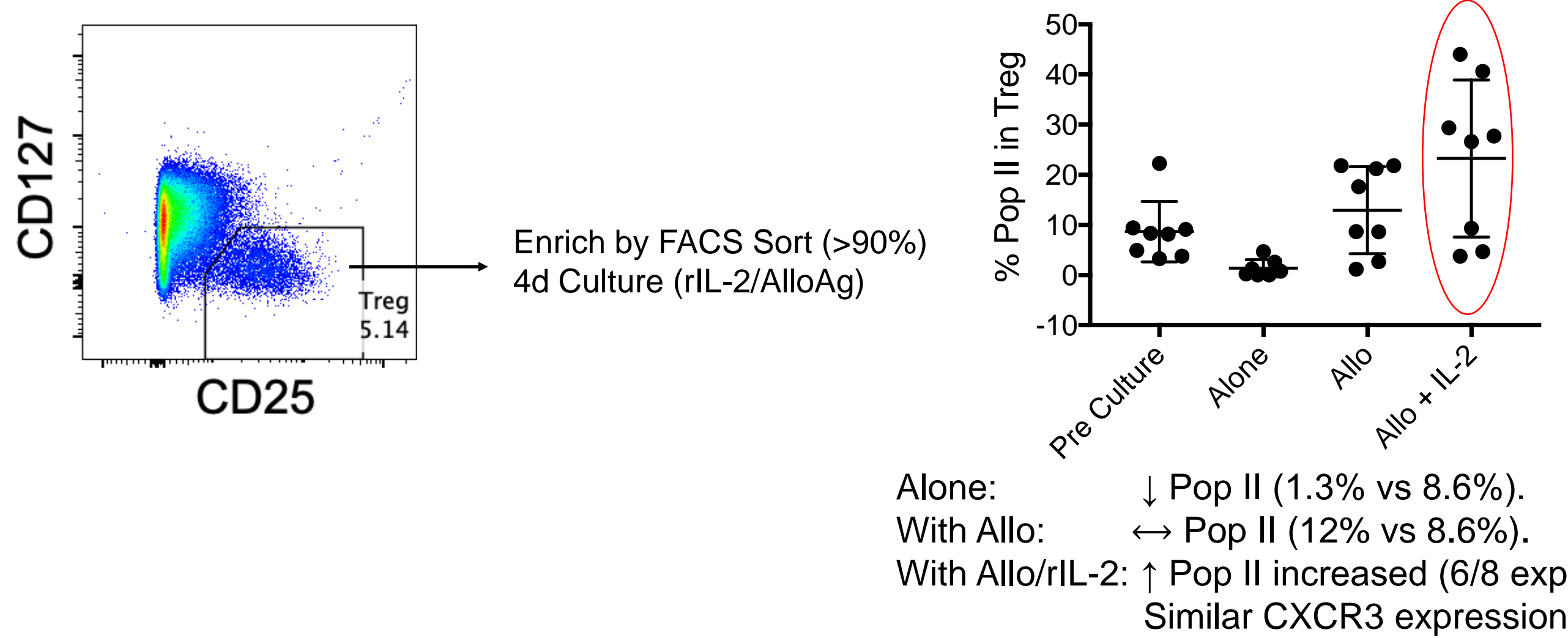
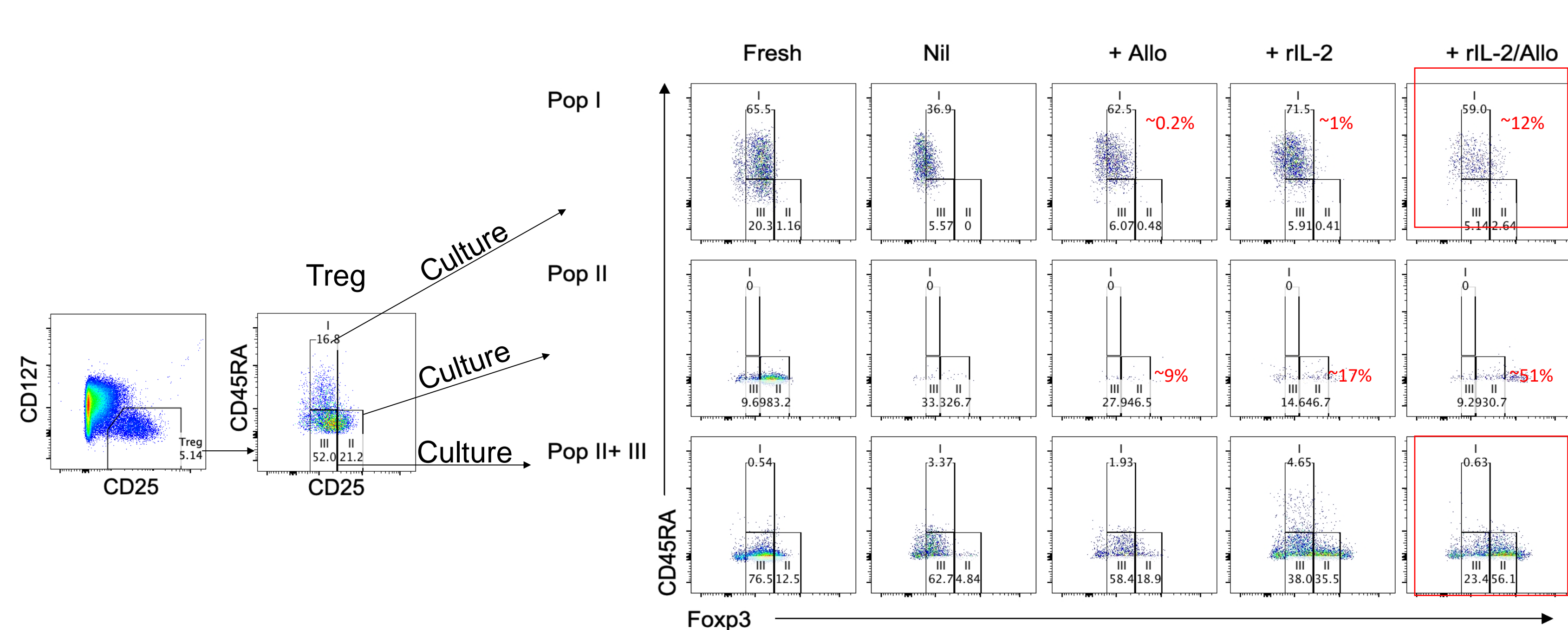
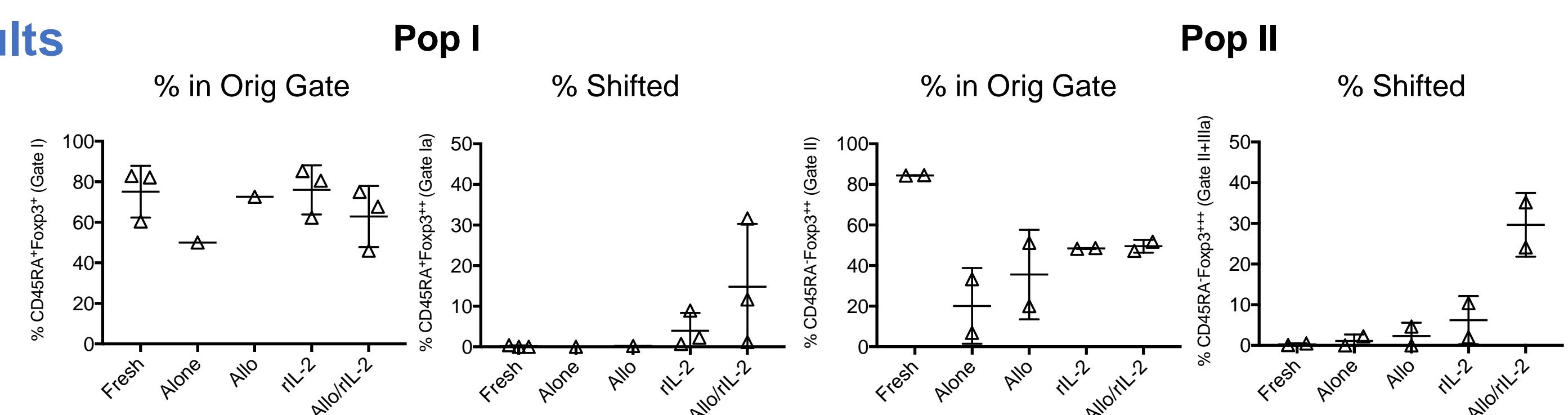


Figure 4. tTreg cultured alone had reduced Pop II compared to the fresh starting population (1.3% vs 8.6%). Culture with alloS preserved Pop II (12% vs 8.6%). IL-2 alone increased Pop II in 5/8 of experiments, similar to the culture with alloS and rIL-2 (6/8 experiments). CXCR3 expression in Pop II was similar in unfractionated Treg to those cultured with IL-2 or alloS alone or IL-2 with alloS (data not shown).

Treg Populations enriched and cultured for 4 days with Allo, rIL-2 or Allo/rIL-2



Pooled results



rIL-2/alloantigen induces expression of IFNGR on Treg

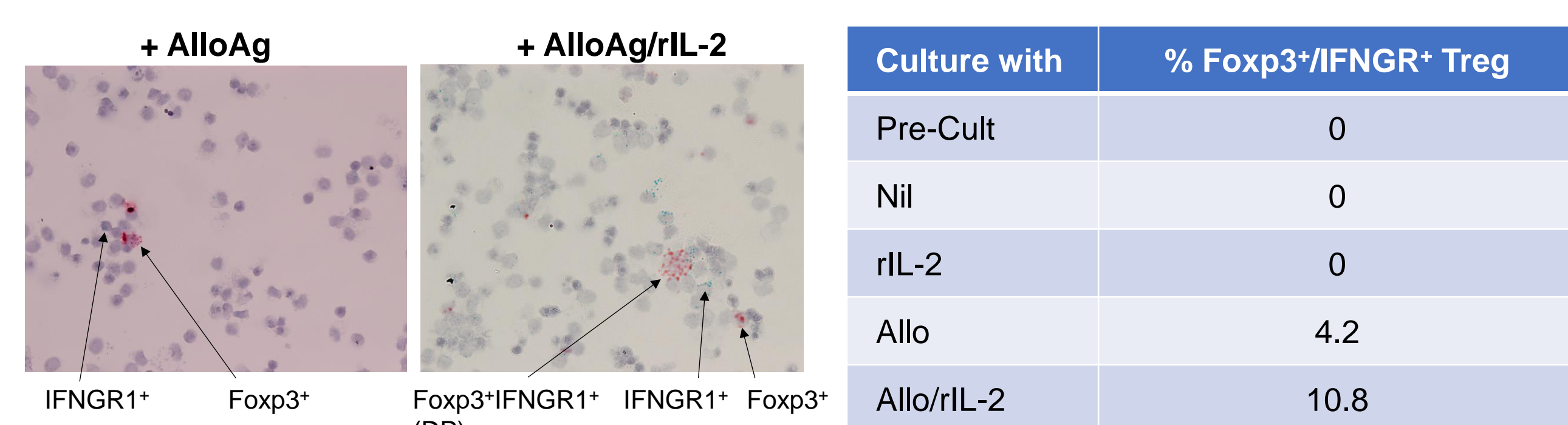


Figure 5. RNAscope showed freshly isolated Treg expressed FOXP3⁺, but not IFNGR1. Treg cultured with alloS and rIL-2 had FOXP3⁺/IFNGR1⁺ double positive cells but also single FOXP3⁺ and IFNGR1⁺ cells. Treg cultured with rIL-2 alone had no IFNGR1⁺ cells whereas Treg cultured with alloS only had single positive FOXP3⁺ and IFNGR1⁺ cells, none were double positive. Treg cultured alone had fewer FOXP3⁺ cells and no IFNGR1⁺ cells.

CONCLUSION

Treg activation can be monitored by changes in expression of CD25, Foxp3, CD45RA and chemokine and cytokine receptors. Human tTreg stimulated with alloS and rIL-2 induced IFNGR, like our rat studies. IFN-γ may induce potent antigen-specific Treg for therapy.

References:

Verma ND, Plain KM, Nomura M, Tran GT, Robinson C, Boyd R, Hodgkinson SJ, Hall BM. CD4⁺CD25⁺ T cells alloactivated ex vivo by IL-2 or IL-4 become potent alloantigen-specific inhibitors of rejection with different phenotypes, suggesting separate pathways of activation by Th1 and Th2 responses. Blood. 2009 Jan 8;113(2):479-87. doi: 10.1182/blood-2008-05-156612. Epub 2008 Sep 30. PMID: 18827184.
Miyara M, Yoshioka Y, Kitoh A, Shima T, Wing K, Niwa A, Parizot C, Taflin C, Heike T, Valeyre D, Mathian A. Functional delineation and differentiation dynamics of human CD4⁺ T cells expressing the FoxP3 transcription factor. Immunity. 2009 Jun 19;30(6):899-911.

ACKNOWLEDGEMENTS: We sincerely appreciate volunteers who generously donated blood for this study. Authors also acknowledge the help provided by nursing staff at the neurology unit, Liverpool hospital and Dr. Jarrad Begg, Cancer Therapy Centre, for helping in cell irradiation.